

Benha University
Faculty of Science
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Carbon Quantum Dots (CQDs) in the targeting and treatment of tumours

Research Project

**Submitted in Partial Fulfilment of the Requirements
for the Award of the Degree of Bachelor of Science
(B.Sc..) in Zoology and Chemistry**

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Acknowledgment

﴿ وَإِذْ تَأْتِيَنَّكُمْ رُبُكُمُ لَئِنْ شَكَرْتُمْ لَأَزِيدَنَّكُمْ ۗ ﴾ سورة إبراهيم – الآية 7

﴿ الْحَمْدُ لِلَّهِ الَّذِي هَدَانَا لِهَذَا وَمَا كُنَّا لِنَهْتَدِيَ لَوْلَا أَنْ هَدَانَا اللَّهُ ﴾ سورة الأعراف – الآية 43

“Gratitude turns what we have into enough.”

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Introduction:

Cancer is one of the common diseases that leads to death in many places due to metastasis. Cancer cells are abnormal cells that grow and divide uncontrollably, invading surrounding tissues and sometimes spreading to all of the body. There are many types of cancer including breast, ovarian, liver, cervical and so on. The mutation and environmental factors like smoking are related to all cancers. The developed countries are more affected than developing countries in this disease.

The most common among women is breast cancer (**Xia *et al.*, 2022**). Breast cancer can lead to a high rate of death due to mutations in genes or other factors (**Mahdavi *et al.*, 2019; Vatankhah *et al.*, 2023; Buist *et al.*, 2018; Hu *et al.*, 2022**). This cancer is related to the same mutation of genes that leads to ovarian cancer (**Hu *et al.*, 2022**). Cervical cancer is more common in developing countries than in developed countries against other types (**Zhang *et al.*, 2020; Prabhu M *et al.*, 2016**). Cancer Therapy by chemicals is more dangerous as it affects normal cells like cancer cells which leads to a high rate of death. Scientists are interested in diagnosing and treating this disease without side effects so they are heading towards nanotechnology.

Nowadays, the whole world is heading towards nanotechnology. CQDs are used as nanoparticles in diagnosis and target treatment. CQDs can penetrate cancer cells easily. Cancer can be diagnosed early by biopsy, computerized tomography scan (CT), magnetic resonance image (MRI), endoscopy and x-rays. However, these tools lead to high costs, lack of awareness and unnecessary treatment. The conventional cancer treatments are surgery, chemotherapy and radiotherapy but there are side effects. There is a need for non-invasive and reliable treatment. CQDs

were used to detect cancer cells by imaging and therapy. So, Nanotechnology has an efficient role in cancer theranostics.

1. Carbon Quantum Dots (CQDs)

1.1 Properties and usage: -

In recent years, Nanotechnology has been used to fight many diseases. Carbon quantum dots, in their nanoscale below 10 nm, are used in many fields, including cell imaging (**Shi *et al.*, 2019**), drug delivery (**Su *et al.*, 2020**), and targeting tumor cells (**Fig. 1**). They are used in cancer therapy due to their properties like biocompatibility, tiny molecules, non-toxicity, and fluorescence. CQDs are synthesised from natural, eco-friendly precursors (**Naik *et al.*, 2022**).

1.2 The precursors for synthesis: -

These precursors are biomolecules or medicinal plants (**Fig. 2**). The biomolecule precursors are proteins and amino acids that are biocompatible and abundant including albumin like bovine serum albumin with formic acid to produce nitrogen-doped CQDs with less toxicity and high fluorescence (**Tan *et al.*, 2015**), carbohydrates like glucose and biomass that contains lipids (**Qiao *et al.*, 2018 and Gusain *et al.*, 2021**), nucleic acid by linking between the two strands of DNA to form DNA-CQDs that lead to carbonization, dehydration, condensation, and polymerisation. The cytosine base in DNA is used to synthesise CQDs by heat at 160°C. These CQDs were used in imaging cells and sensing due to their photostability, biocompatibility, and wide-range pH (**Luo *et al.*, 2018**).

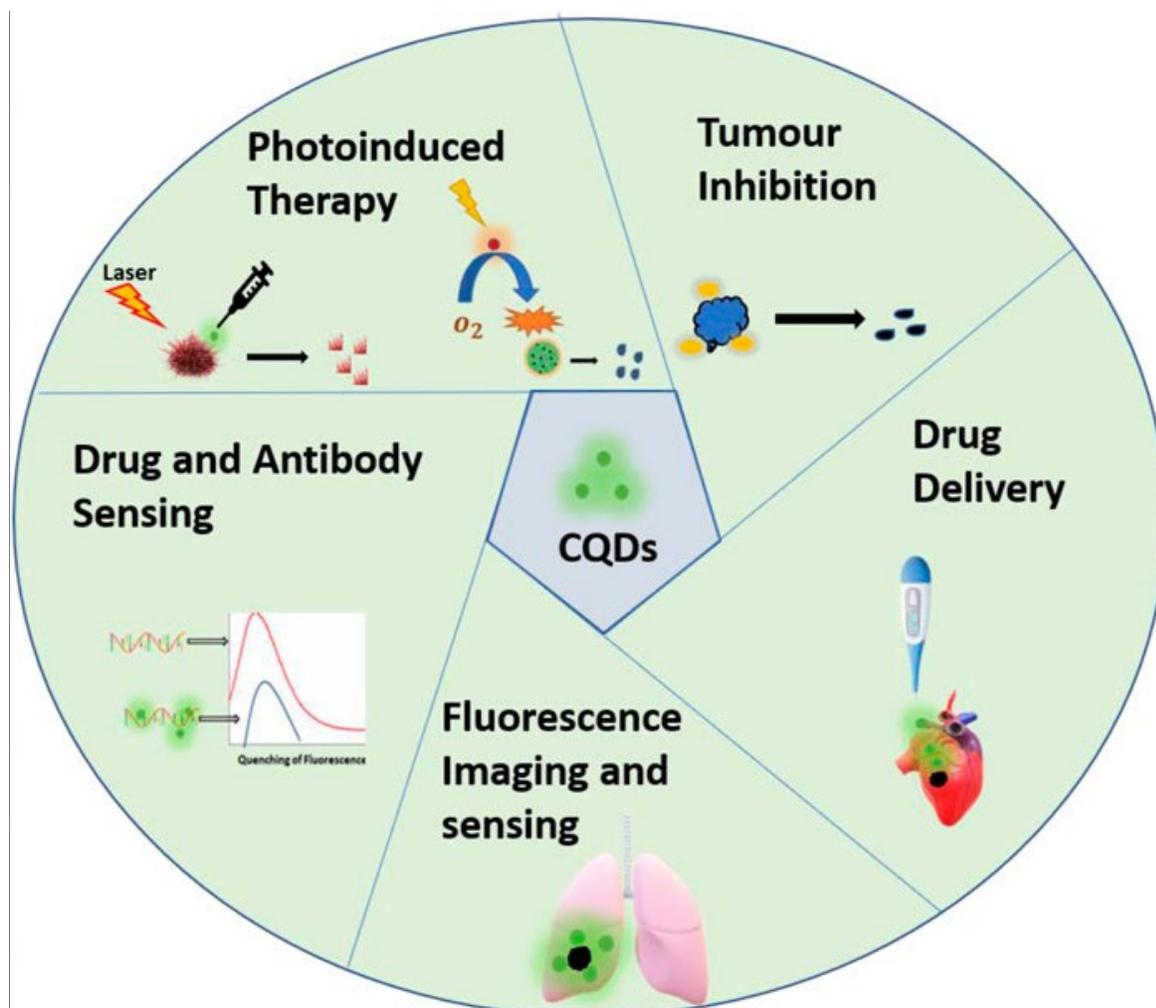


Figure 1: CQDs in cancer diagnosis and treatment. (Naik *et al.*,2021)

In the last studies, medicinal plant-based CQDs have drawn attention (Naik *et al.*,2021) due to their medicinal properties, such as anticancer properties, which make them useful in cancer treatment (Park *et al.*, 2017). These CQDs are efficient in this field due to their photostability, high photoluminescence (PL), water solubility and nontoxicity (Naik *et al.*,2021). Some of these plants used to synthesise CQDs are green tea (He *et al.*, 2021), Ginger juice (Naik *et al.*,2022), and walnut oil (Arkan *et al.*, 2018).

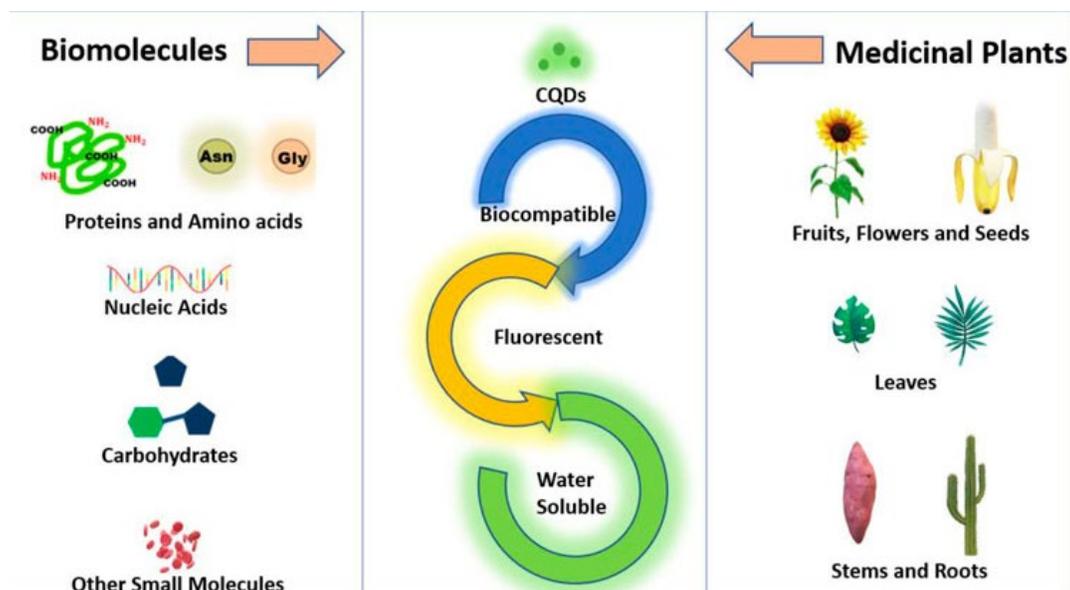


Figure 2: The synthesis of CQDs from biomolecules and medicinal plants. (Naik *et al.*, 2021)

1. Breast Cancer

Breast cancer has become the main reason women die (Obeagu *et al.*, 2024). Although BC is an issue for global public health, China ranks first globally due to its large population and high cancer rate (Xia *et al.*, 2022). BC can be determined by a nipple discharge, breast shape or size change, or the discovery of breast lumps or mastalgia, even though breast examination is essential (Heer *et al.*, 2020). It is from a mutation in breast cells. It has some types, including ductal, lobular (Edward *et al.*, 2021 and Obeagu *et al.*, 2023), triple-negative breast cancer, breast Paget disease, inflammatory breast cancer, and soft tissue sarcoma (Jagsi *et al.*, 2022). The most common cancers are ductal and lobular, which are present in location and still in situ or spread in adjacent tissue (Iatrakis *et al.*, 2021).

BC is a multifactorial disease that comes with some risk factors, including diet, obesity, genetics, smoking, drinking, and cosmetics that contain estrogen

(**Mahdavi et al., 2019**). Despite this, breast cancer can develop without any risks in women than men because of exposure to estrogen and progesterone. According to this, late menopause leads to BC due to exposure to estrogen and progesterone (**Vatankhah et al., 2023**). The ductal or lobular carcinoma in situ can appear again in women due to their personal history, which leads to increasing the risk of this disease (**Buist et al., 2018**). The increasing number of injuries by BC in close blood relations is related to family history (**Maio et al., 2021**). Mutations in BRCA genes, including BRCA1 & BRCA2, lead to uncontrolled growth in BCs and tumor cells. The presence of 1 or 2 copies of the mutated gene leads to a 50 % chance of inheriting in the family, and these genes can also lead to ovarian cancer (**Hu et al., 2022**). Early pregnancy decreases the rate of chance injury by BC (**Garnæs et al., 2022**).

2.1 The CQDs in targeting tumor cells:

Methotrexate, MTX, is a drug used as an anticancer but non-targeting for tumor cells. That leads to some side effects in the body (**Zuber et al., 2021 and Poursadegh et al., 2024**). FA-CQDs with Ex sourced from BC cells were used as targeted MTX to avoid that (**Yang et al., 2023 and Tiwari et al., 2024**). Ex derived from cancer cells and used as a targeted drug in a range from 40 to 100 nm in size due to their membrane composition, bioavailability, and diminished off-target cytotoxicity (**Gilligan et al., 2017 and Tiwari et al., 2023**) and these vesicles don't affect normal cells (**Saw et al., 2019 and Zhu et al., 2021**). There are folate receptors in the malignant cells (**Prieto et al., 2020 and Khoshnood et al., 2023**) that lead to using it in treating cancer cells when attached to CQDs (**Sattariazar et al., 2023 and Zahed et al., 2024**).

2.2 Drug loading and cell viability

Drug loading and cellular uptake study using CQDs, MTX-CQDs and Ex@MTX-CQDs in cancer cells (Fig. 3).

The release of MTX from Ex@MTX-CQDs at conditions pH 5 and 7.4 indicates that 65% of MTX was released in acidic conditions like the tumor microenvironment, and around 10% was released in neutral environments like normal cells. That leads to low toxicity in normal cells and high targeting (Fig. 4 A).

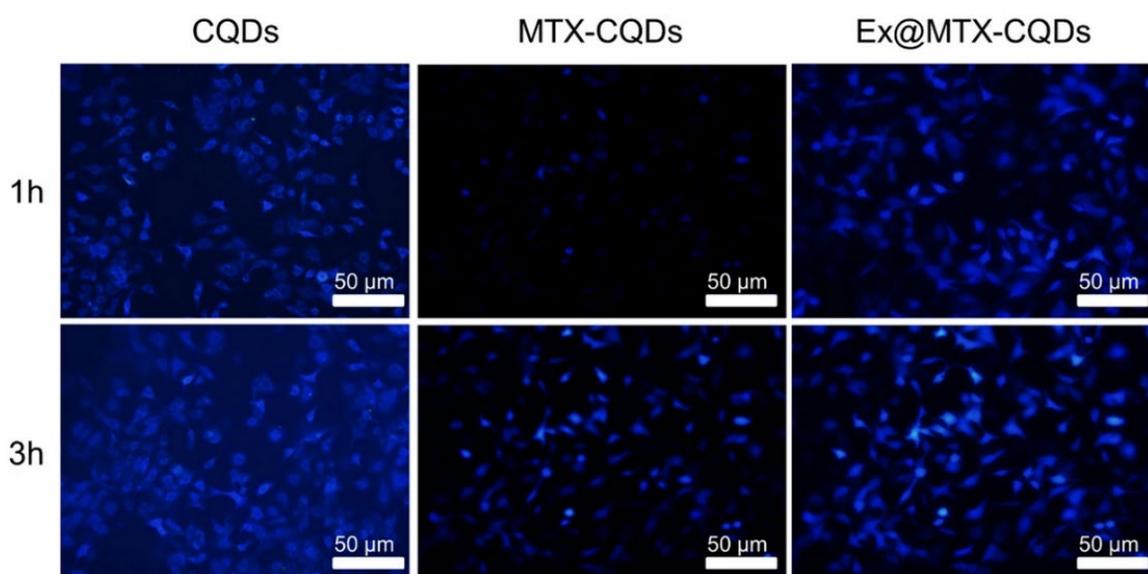


Figure 3: The cellular uptake images of CQDs, MTX-CQDs, and Ex@MTX-CQDs. (Kazeminava *et al.*, 2024)

The MTT assay measured the cell viability in the MCF-7 cell line. The effect of Ex@MTX-CQDs leads to less toxicity against breast cancer than CQDs, MTX, Ex, or MTX-CQDs. Ex@MTX-CQDs marked cytotoxicity against the MCF-7 cell line (Fig. 4 B).

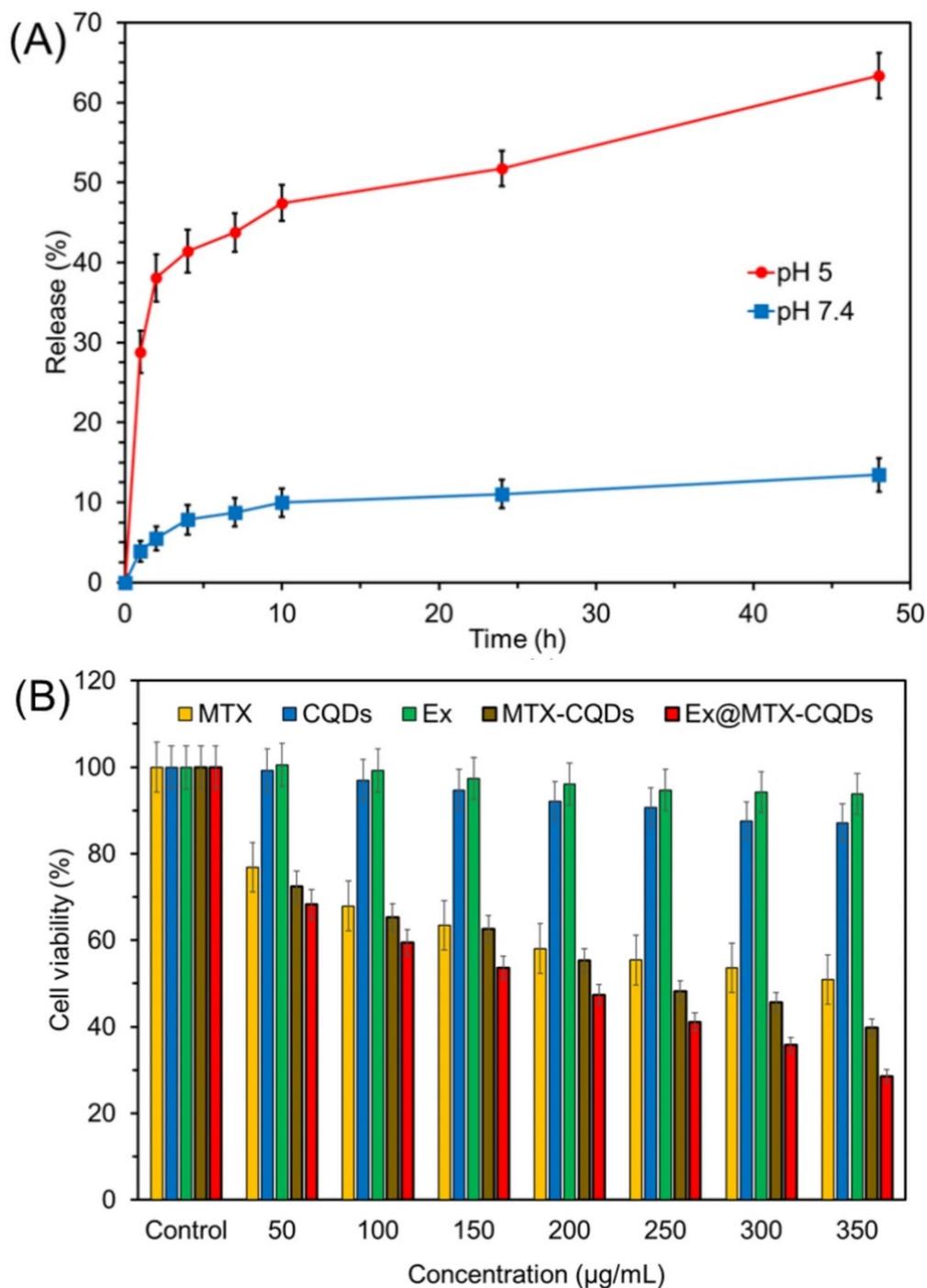


Figure 4: MTX release profile of Ex@MTX-CQDs (A) and the cell viability study of MCF-7 cell after incubation at various concentrations (mg/mL) of samples 48 h (B). (Kazeminava *et al.*, 2024)

2. Ovarian Cancer

Ovarian cancer is a lethal disease that leads women to die at an older age (Feeney *et al.*, 2020 and Siegel *et al.*, 2020). In the 21st century, the incidence of ovarian cancer has decreased in Northern America and Western Europe (Siegel *et al.*, 2020). Despite the decreasing incidence rate, ovarian cancer is the most dangerous and leads to 2-thirds of deaths because of being diagnosed at later stages (Aus *et al.*, 2020). The WHO has classed ovarian cancer into five major types: mucinous carcinoma, endometrioid carcinoma, high-grade serous carcinoma, low-grade carcinoma, and clear cell carcinoma due to differences in cell origin (Kobel *et al.*, 2022). Women between 60 and 65 years old have the highest rate of ovarian cancer in developed countries (Siegel *et al.*, 2020). The rate of ovarian cancer has increased because of the Western lifestyle, family size, feeding babies milk formula and a decrease in the rate of pregnancy (Ali *et al.*, 2018). The survival rate has decreased at advanced stages like (III, IV) (Timmermans *et al.*, 2018) and varies due to different disease histotypes (Zhou *et al.*, 2021). An earlier diagnosis of breast cancer leads to an earlier diagnosis of ovarian cancer (Ali *et al.*, 2023).

Ovarian cancer includes epithelial tumors, germ cell tumors and sex cord stromal tumors that are heterogeneous histologic types. When 10-15 % of ovarian cancer is due to genetics, there are multiple factors because of the complex and heterogeneous epithelial cells (Janardhan *et al.*, 2015). Surface epithelial cells lead to 90% of ovarian cancer (Ali *et al.*, 2023). Developed countries have a higher rate of ovarian cancer than developing countries because of increasing life expectancy, decreasing fertility rate, feeding babies in milk formula, western lifestyle, and increased daily intake of fatty diet and dense caloric food (Rice *et al.*, 2020). Around 75% of ovarian cancer in older women is diagnosed after menopause (Bandera *et al.*, 2016) that's because of poor prognosis (Ali *et al.*, 2020).

Ovarian cancer affects white American women more than African American women (**Bandera *et al.*, 2016**) because of BRCA mutations. Although BRCA1 and BRCA2 have the same functions in DNA repair, they affect cancer differently by their mutations. BRCA1 mutations are affected two to threefold compared with BRCA2 mutations. The injured first or second-degree relatives lead to BRCA1 & 2 mutations that affected ovarian cancer by 84% from BRCA1 mutations and 16% from BRCA2 mutations (**Ali *et al.*, 2023**). A recent study shows that BRCA1 mutations are more aggressive than BRCA2 mutations because they lead to reprogramming in tumor cells (**Bruand *et al.*, 2021**).

3.1 The CQDs in targeting tumor cells:

CQDs are highly toxic to tumor cells. Copper metal is used to treat SKOV3 cells. Cu-based nanomaterials affect cell cycle regulation more than other nanomaterials related to carcinogenic processes (**Seong *et al.*, 2015 and Chen *et al.*, 2021**).

The CQDs/Cu₂O have higher toxicity against SKOV3 cells than CQDs or Cu₂O, and the copper has high stability (**Chen *et al.*, 2021**). The green nucleus indicates the controlled healthy cell, the orange nucleus indicates the late stage of apoptosis and the necrotic cells with uniformly red nuclei (**fig .5**).

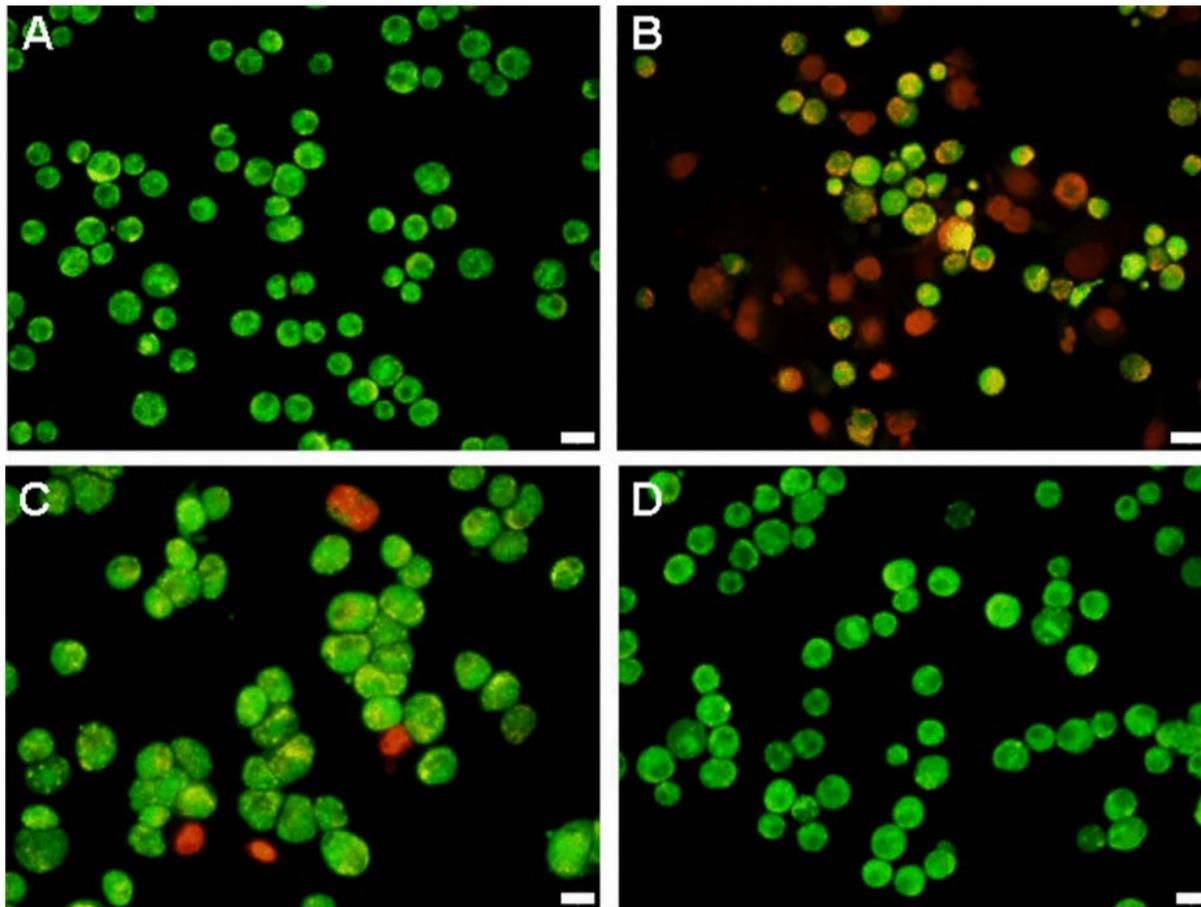


Figure 5: Fluorescence photomicrographs changes by AO/EB staining of SKOV3 cells after treatment with CQDs/Cu₂O (B), CQDs (C), or Cu₂O (D) for 24 h compared to controls (A), respectively. Scale bar 20 μ m. (Chen *et al.*, 2021)

The CQDs/Cu₂O also have higher toxicity in cancer cells than OXA and ART. The IC₅₀ of CQDs/Cu₂O in SKOV3 cells is less than the IC₅₀ of each of OXA and ART (WangJ *et al.*, 2016).

3.2 The effect of CQDs/Cu₂O:

The effect of CQDs in SKOV3 cells was non-toxic and biocompatible (Shereema *et al.*, 2015). In the MTT assay, a dependent concentration of CQDs/Cu₂O led to cytotoxicity and the SKOV3 cells had a higher inhibition rate between other cancer

cell lines like (Hela, A549, HT-29, HCT116, BABL-3T3, HEK293T, J224AI) that indicate the CQDs/Cu₂O lead to inhibit the growth of SKOV3 cells (fig. 6).

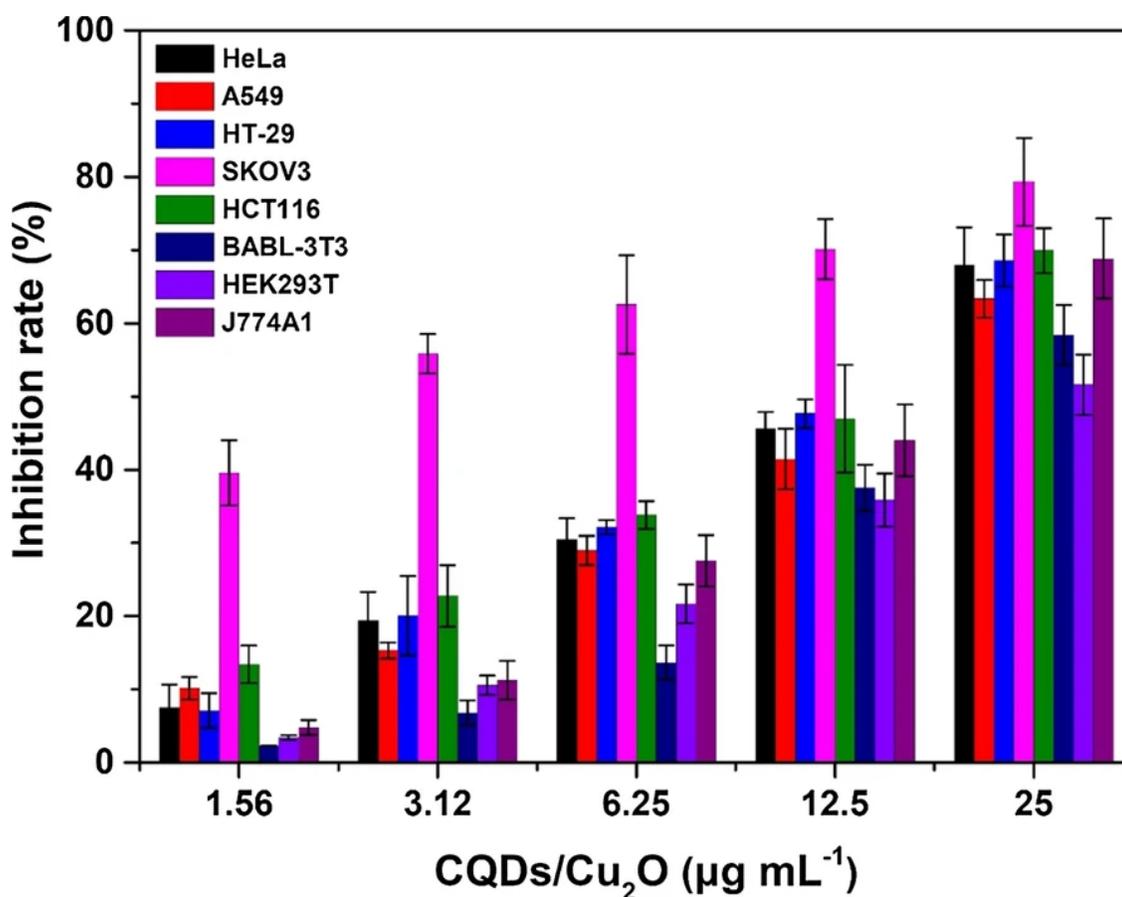


Figure 6: Differential cytotoxicity of CQDs/Cu₂O in cancer cells (HeLa, A549, HT-29, SKOV3, HCT116) and normal cells (BABL-3T3, HEK293T, J774A1) by the MTT assay for 24 h. (Chen *et al.*, 2021)

The WST assay shows the results from the MTT assay by the same range of inhibitory concentration. A low concentration of the CQDs/Cu₂O leads to the differentiation between SKOV3 cancer and other cells associated with anticancer drugs (WangJ *et al.*, 2016).

3. Liver cancer

Liver cirrhosis is a dangerous disease that spreads worldwide. The hepatic stellate cells have a role in this field and can develop cancer. It can be developed by the potential polarization of CAFs (cancer-associated fibroblasts), TAMs (tumor-associated macrophages) and TECs (tumor endothelial cells) (Ma *et al.*, 2019). There are two types of liver cancer (Hepatocyte cancer and Cholangiocyte cancer), and they lead to differences in histology and molecular features. Features of HCCs are vascular and stromal invasion, thick cell trabeculae, increased cell density, loss of the reticulin framework, and a pseudo-glandular pattern (Cogliati *et al.*, 2023). The iCCA differentiates according to the affected bile duct (Vijgen *et al.*, 2017 and Kendall *et al.*, 2019). The HCC contains two phenotype classes: proliferation and non-proliferation (Zucman-Rossi *et al.*, 2015 and Ally *et al.*, 2017), and the iCCA includes proliferation and inflammation (Cogliati *et al.*, 2023). The HCC has unique molecular and histological features in the proliferation that lead to HBV infection, poor differentiation, pro-proliferative pathways, a worse prognosis, and increased rate of TP53 mutations, and the non-proliferation leads to a better outcome and WNT signalling due to CTNNB1 mutations (Ally *et al.*, 2017). iCCA is similar to HCC and leads to common IDH1/2 mutations and FGFR2 fusions (Sia *et al.*, 2015 and Nakamura *et al.*, 2015).

HBV and HCV can induce immunity to promote or suppress carcinogenesis. HBV can inhibit the effect of T-cells on cancer cells as the T_{reg} is highly immunosuppressive (Pallett *et al.*, 2015; Tan *et al.*, 2019 and Lim *et al.*, 2019). HCV can promote cancer cells by loss of IL-2 secreting CD4⁺ T-helper cells and the presence of exhaustion markers like TIM3, PD1 and galectin 9 that secrete from monocytes and promote T_{reg} cell expansion (Hofmann *et al.*, 2021). During NASH, there is a crosstalk between immune cells and hepatocytes that leads to the

development of cancer like CD8⁺PD1⁺T that induces liver damage in preclinical mouse models because of metabolic stimuli and CD⁺IL-17A⁺T helper 17 cells that induces white adipose tissue neutrophil infiltration that mediates insulin resistance and fatty acid release (Gomes *et al.*, 2016; Ma *et al.*, 2016; Pfister *et al.*, 2021 and Dudek *et al.*, 2021).

Cancer-associated fibroblasts (CAFs) can promote tumor cells by ligand-receptor interactions, deposition of ECM components and release of growth factors and inflammatory cytokines (Sahai *et al.*, 2020; Donne *et al.*, 2023 and Llovet *et al.*, 2023). CAFs can mediate the secretion of collagen, proteoglycans and fibronectins with the crosslinking enzymes that promote tumor cell growth and increase tissue stiffness that activates integrins, SRC family kinases, FAK and YAP-TAZ signaling that leads to increased cancer cells by its pro-migratory and pro-proliferative in both cancer and stromal cells (Cogliati *et al.*, 2023). For example, YAP-TAZ signaling is used as an anti-apoptotic protein in cancer cells, leading to tumor proliferation (Stein *et al.*, 2015 and Chang *et al.*, 2015).

4.1 CQDs and Huh7 cell line:

The CQDs were used with tryptophan and sorbitol for high stability and optical properties. The TC-WS-CQDs lead to cell toxicity up to a certain concentration, which leads to targeting tumor cells. The B -, G -, and R-WS-CQDs indicated the endocytosis of the TC-WS-CQDs in tumor cells after incubation. The nystatin and chlorpromazine incubate the CQDs to detect the location of tumor cells (Fig. 7).

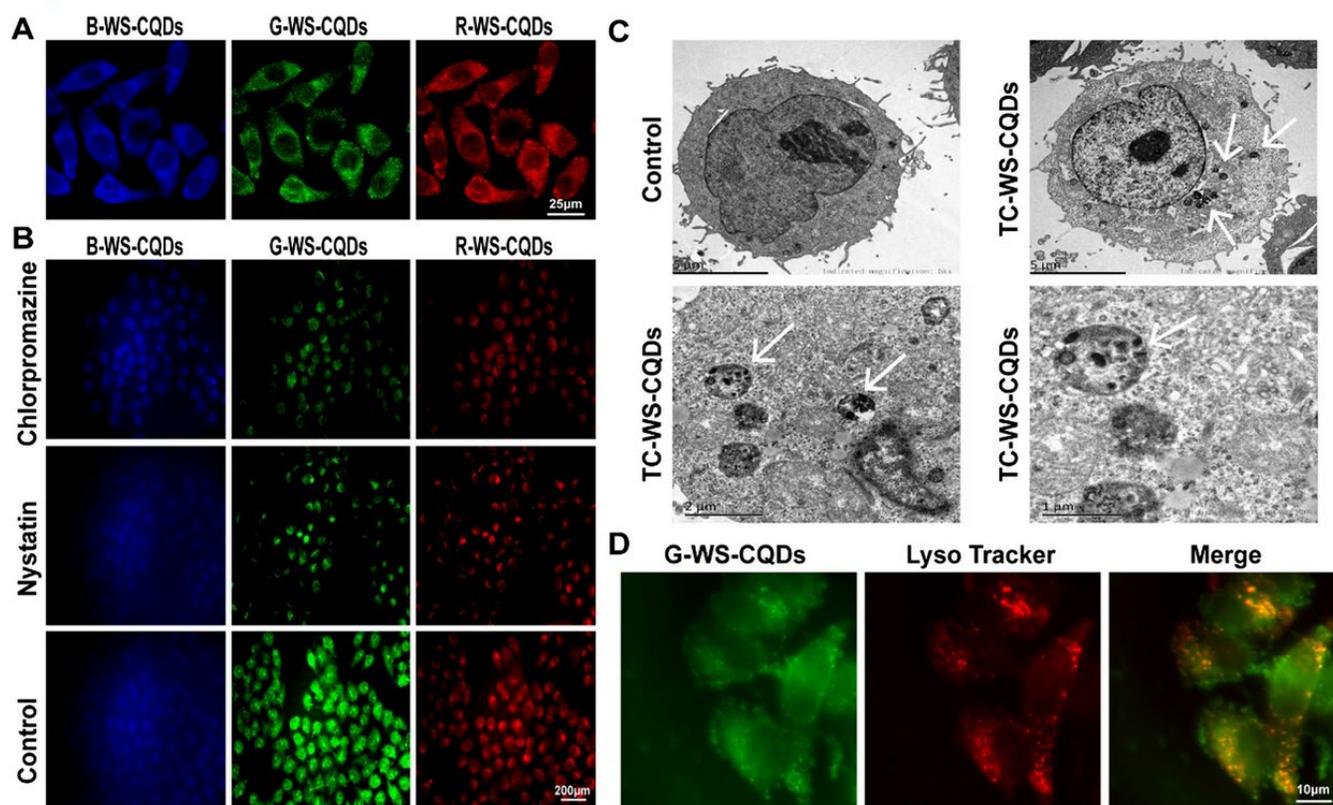


Figure 7: (A) LSCM images of Huh7 cells with TC-WS-CQDs incubation for 6 h. (B) Huh7 cells were treated with TC-WS-CQDs alone and co-treated with Nystatin (30 μM) or Chlorpromazine (20 μM) for 6 h. (C) TEM assessed the intracellular localization of TC-WS-CQDs in Huh7 cells. Arrows indicate intracellular vesicles engulfing TC-WS-CQDs in the cytoplasm. (D) G-WS-CQDs co-localise with lysosomes as tracked by LysoTracker. (Wang *et al.*, 2022)

4.2 Treating by CQDs:

The CQDs induce high radiation therapy, decreasing the tumour cells' weight but not affecting the healthy cells (Fig. 8).

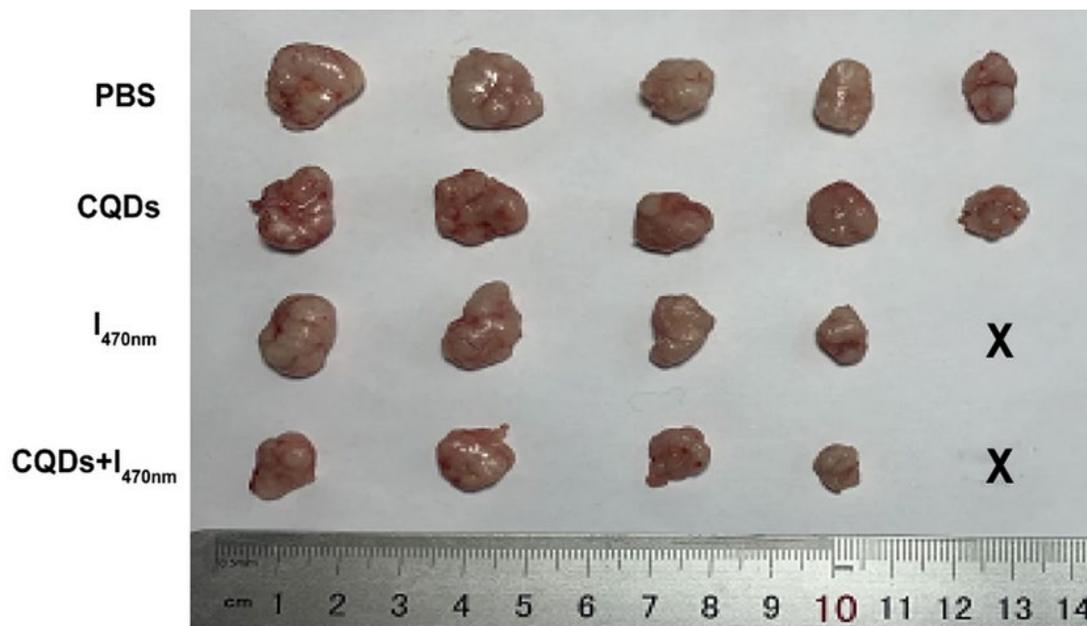


Figure 8: The effect of CQDs, I_{470nm} and CQDs+ I_{470nm} on the size of tumor cells. (Wang *et al.*, 2022)

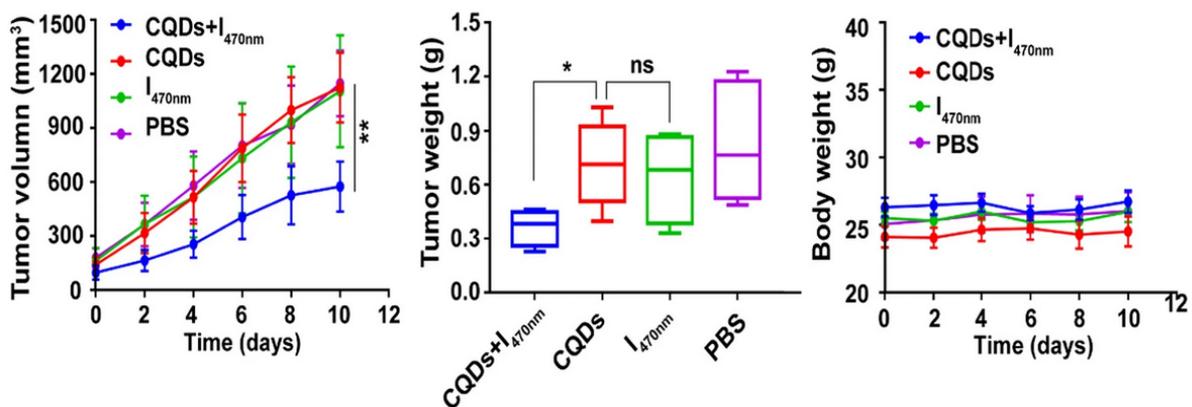


Figure 9: The effect of CQDs, I_{470nm} and CQDs+ I_{470nm} on the tumor volume (mm³), tumor weight (g) and body weight (g). (Wang *et al.*, 2022)

4. Cervical cancer

Cervical cancer is the second most common cancer (**Zhang *et al.*, 2020**) that leads to death (**Mattiuzzi *et al.*, 2020**). This cancer has a higher rate in developing countries than in developed countries (**Zhang *et al.*, 2020**). The low and middle-income individuals in developing countries have a higher rate of death from this cancer than in wealthier countries (**Prabhu *et al.*, 2016**). The percentage of infection rate varies according to HPV genotype (**Zhang *et al.*, 2020**). Women with HPV have a high risk of incidence of cervical intraepithelial neoplasia (CIN) and HPV types (**Stelzle *et al.*, 2021 and Yuan *et al.*, 2021**).

HPV have many genotypes, there are around 13 of these that lead to cervical cancer (**Perkins *et al.*, 2023**). The percentage of cancer changes according to the genotype of HPV that is associated with alpha-9 and so on (**Table. 1**). The high-risk or oncogenic HPV types, including HPV16 and HPV18, can lead to cervical cancer (**Cohen *et al.*, 2019**). Sexual behaviour affects the incidence of HPV. HPV can spread after sexual intercourse, leading to cervical intraepithelial neoplasia (CIN) (**Zhang *et al.*, 2020**).

Table 1: Indicate the percentage of cancer cells according to the genotype of HPV. (Bedell *et al.*, 2020)

Carcinogenic human papillomavirus (HPV) type	Proportion of cervical cancers, %	9-Year risk of progression of incident HPV infection to cervical intraepithelial neoplasia grade 3 or worse (CIN3+)	HPV species	Risk group	Included in 9-valent vaccine
16	60.3	6.3	Alpha-9	Highest	Yes
18	10.5	3.0	Alpha-7	High	Yes
45	6.1	2.2	Alpha-7	High	Yes
33	3.7	4.5	Alpha-9	Medium	Yes
31	3.6	2.2	Alpha-9	Medium	Yes
52	2.7	2.2	Alpha-9	Medium	Yes
58	2.2	1.9	Alpha-9	Medium	Yes
35	2.0	2.8	Alpha-9	Medium	No
39	1.6	1.1	Alpha-7	Lower	No
51	1.2	1.1	Alpha-5	Lower	No
59	1.1	0.9	Alpha-7	Lower	No
56	0.9	0.8	Alpha-6	Lower	No
68	0.6	1.0	Alpha-7	Lower	No

Multiple sexual partners and early age at first intercourse cause a high risk of cervical cancer (Liu *et al.*, 2015). Oral contraceptive pills (OC) are associated with cervical cancer as the increase in the use of OC leads to an increase in the risk of cervical cancer (Zhang *et al.*, 2020).

5.1 CQDs against cervical cancer:

Nowadays, cervical cancer has spread among females in low and middle-income nations (Bentivegna *et al.*, 2016). The World Health Organization (WHO) reports that cervical cancer is the fourth cancer spread among women (Alam *et al.*, 2022). CQDs face this cancer because of their properties, including photostability, low toxicity and biocompatibility (Singh *et al.*, 2018). CQDs are synthesised from chlorophyll to be non-toxic and don't negatively affect the body (Devi *et al.*, 2019 and Unnikrishnan *et al.*, 2020). Chl-CQDs were used in the diagnostics and

therapeutics of tumor cells due to their long absorption wavelength (Wu *et al.*, 2021).

5.2 ChI-CQDs suppress the cell viability of SiHa cells:

MTT assay indicates that the ChI-CQDs cause inhibition in SiHa cancer and do not affect the normal kidney cells HEK-293. The viability of SiHa cells decreased by increasing the concentration of the drug. (fig. 10)

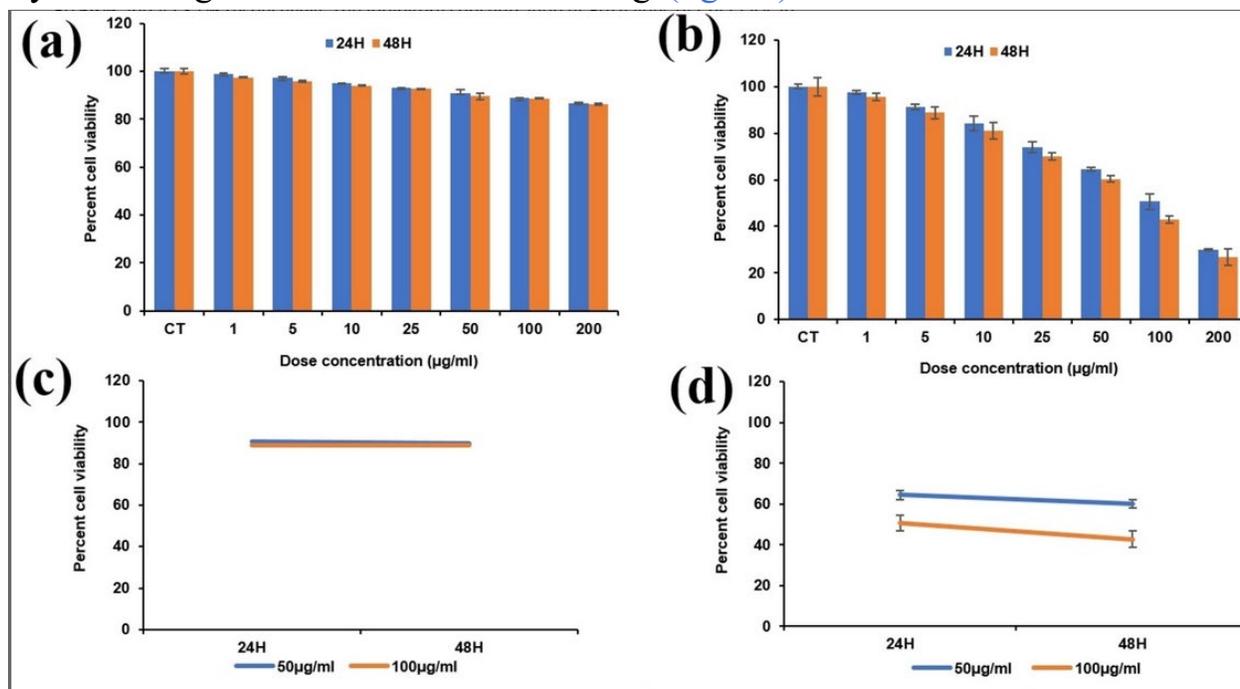


Figure 10: The effect of ChI-CQDs on the proliferation of cervical cancer cell line. HEK-293 and SiHa cells were exposed to indicated concentrations of doses and the cell viability was assayed using MTT as a substrate by taking absorbance at 570 nm. (Alam *et al.*, 2022)

5.3 ChI-CQDs promote morphological changes in SiHa cells:

ChI-CQDs lead to uneven form, cell shrinkage and separation from the surface in SiHa cells. CQDs also lead to SiHa cell apoptosis, which was indicated by DAPI staining or by using AOPI staining. CQDs show the SiHa cells died by red

fluorescence, and the non-died or normal cells of the kidney appear with green fluorescence (Fig. 11).

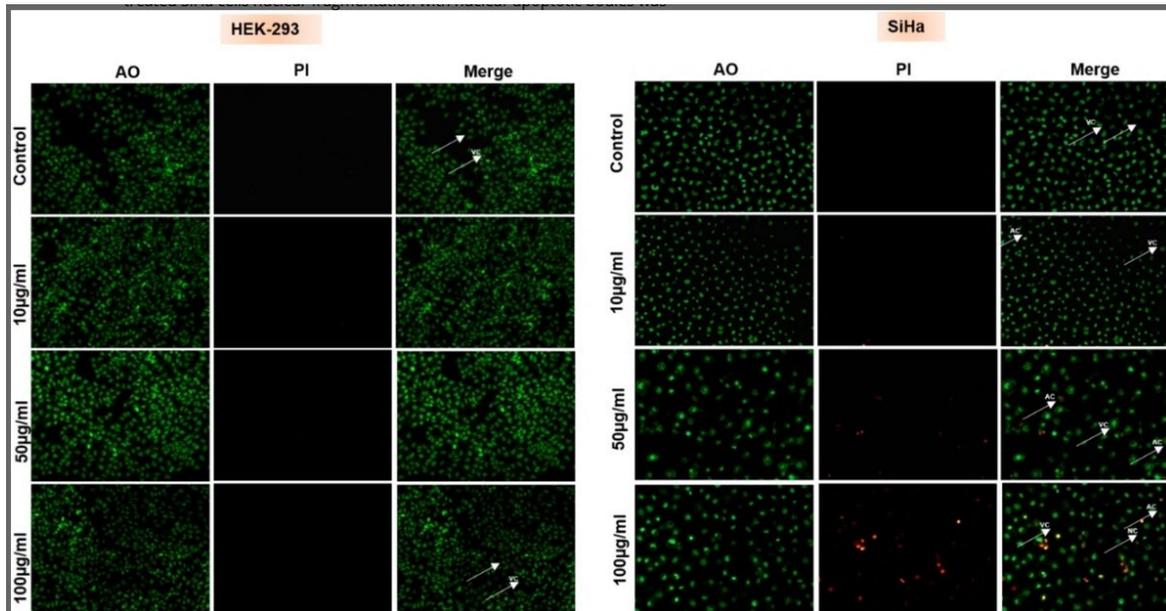


Figure 11: The effect of AOPI on SiHa cell and HEK-293 cell. (Alam *et al.*, 2022)

ChI-CQDs lead to the damage plasma membrane in tumor cells and don't affect normal cells and that was indicated by LDH release from the plasma membrane (Fig. 12).

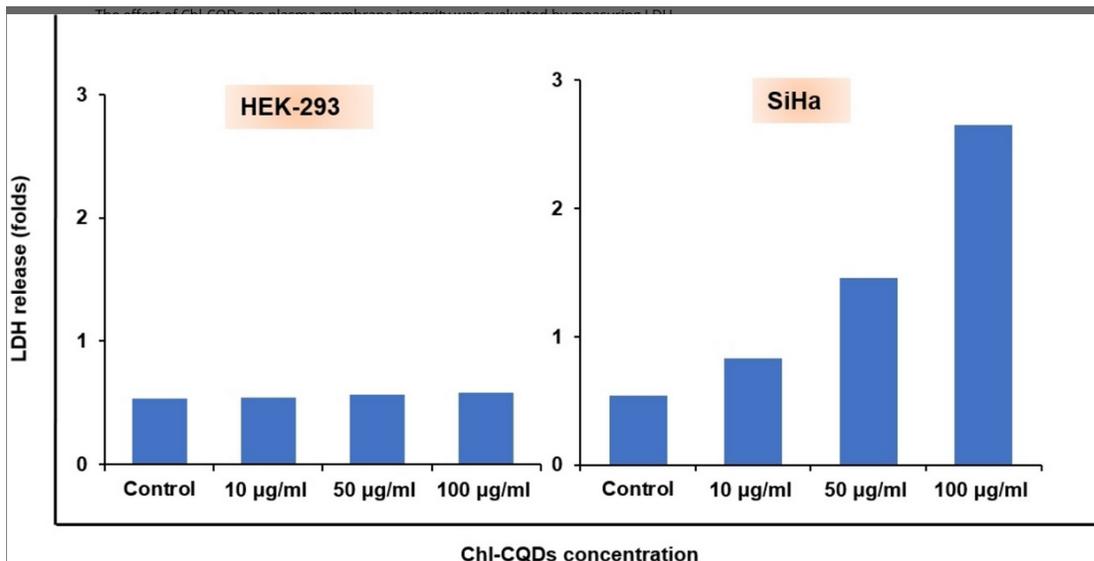


Figure 12: The effect of concentration of ChI-CQDs on releasing LDH (folds). (Alam *et al.*, 2022)

References

1. Alam, M. B., Minocha, T., Yadav, S. K., & Parmar, A. S. (2022). Therapeutic potential of chlorophyll functionalized carbon quantum dots against cervical cancer. *ChemistrySelect*, 7(48), e202204562.
2. Ali AT. Towards prevention of ovarian cancer. *Curr Cancer Drug Target* 2018; 18: 522-537.
3. Ali, A. T., Al-Ani, O., & Al-Ani, F. (2023). Epidemiology and risk factors for ovarian cancer. *Menopause Review/Przegląd Menopauzalny*, 22(2), 93-104.
4. Ally, A., Balasundaram, M., Carlsen, R., Chuah, E., Clarke, A., Dhalla, N., ... & Ferguson, M. L. (2017). Comprehensive and integrative genomic characterization of hepatocellular carcinoma. *Cell*, 169(7), 1327-1341.
5. Aus, A. T. (2020). Can we prevent ovarian cancer?. *Ceska Gynekologie*, 85(1), 49-58.
6. Bandera, E. V., Lee, V. S., Rodriguez-Rodriguez, L., Powell, C. B., & Kushi, L. H. (2016). Racial/ethnic disparities in ovarian cancer treatment and survival. *Clinical cancer research*, 22(23), 5909-5914.
7. Bedell, S. L., Goldstein, L. S., Goldstein, A. R., & Goldstein, A. T. (2020). Cervical cancer screening: past, present, and future. *Sexual medicine reviews*, 8(1), 28-37.
8. Bentivegna, E., Gouy, S., Maulard, A., Chargari, C., Leary, A., & Morice, P. (2016). Oncological outcomes after fertility-sparing surgery for cervical cancer: a systematic review. *The Lancet Oncology*, 17(6), e240-e253.
9. Bruand, M., Barras, D., Mina, M., Ghisoni, E., Morotti, M., Lanitis, E., ... & Coukos, G. (2021). Cell-autonomous inflammation of BRCA1-deficient ovarian cancers drives both tumor-intrinsic immunoreactivity and immune resistance via STING. *Cell reports*, 36(3).

10. Buist, D. S., Abraham, L., Lee, C. I., Lee, J. M., Lehman, C., O'Meara, E. S., ... & Breast Cancer Surveillance Consortium. (2018). Breast biopsy intensity and findings following breast cancer screening in women with and without a personal history of breast cancer. *JAMA internal medicine*, 178(4), 458-468.
11. Chang, C., Goel, H. L., Gao, H., Pursell, B., Shultz, L. D., Greiner, D. L., ... & Mercurio, A. M. (2015). A laminin 511 matrix is regulated by TAZ and functions as the ligand for the $\alpha 6\beta 1$ integrin to sustain breast cancer stem cells. *Genes & development*, 29(1), 1-6.
12. Chen, D., Li, B., Lei, T., Na, D., Nie, M., Yang, Y., ... & Wang, J. (2021). Selective mediation of ovarian cancer SKOV3 cells death by pristine carbon quantum dots/Cu₂O composite through targeting matrix metalloproteinases, angiogenic cytokines and cytoskeleton. *Journal of Nanobiotechnology*, 19, 1-17.
13. Cogliati, B., Yashaswini, C. N., Wang, S., Sia, D., & Friedman, S. L. (2023). Friend or foe? The elusive role of hepatic stellate cells in liver cancer. *Nature Reviews Gastroenterology & Hepatology*, 20(10), 647-661.
14. Cohen, P. A., Jhingran, A., Oaknin, A., & Denny, L. (2019). Cervical cancer. *The Lancet*, 393(10167), 169-182.
15. Devi, P., Saini, S., & Kim, K. H. (2019). The advanced role of carbon quantum dots in nanomedical applications. *Biosensors and Bioelectronics*, 141, 111158.
16. Donne, R., & Lujambio, A. (2023). The liver cancer immune microenvironment: Therapeutic implications for hepatocellular carcinoma. *Hepatology*, 77(5), 1773-1796.
17. Dudek, M., Pfister, D., Donakonda, S., Filpe, P., Schneider, A., Laschinger, M., ... & Knolle, P. A. (2021). Auto-aggressive CXCR6⁺ CD8 T cells cause liver immune pathology in NASH. *Nature*, 592(7854), 444-449.

18. Edward, A., Obeagu, E. I., Okorie, H. M., Vincent, C. C. N., & Bot, Y. S. (2021). Studies of serum calcium, inorganic phosphate and magnesium levels in lactating mothers in Owerri. *J Pharm Res Int*, 33(41B), 209-216.
19. Feeney, L., Harley, I. J., McCluggage, W. G., Mullan, P. B., & Beirne, J. P. (2020). Liquid biopsy in ovarian cancer: Catching the silent killer before it strikes. *World Journal of Clinical Oncology*, 11(11), 868.
20. Gilligan, K. E., & Dwyer, R. M. (2017). Engineering exosomes for cancer therapy. *International journal of molecular sciences*, 18(6), 1122.
21. Gomes, A. L., Teijeiro, A., Burén, S., Tummala, K. S., Yilmaz, M., Waisman, A., ... & Djouder, N. (2016). Metabolic inflammation-associated IL-17A causes non-alcoholic steatohepatitis and hepatocellular carcinoma. *Cancer cell*, 30(1), 161-175.
22. Gusain, D., Renuka, N., Guldhe, A., & Bux, F. (2021). Use of microalgal lipids and carbohydrates for the synthesis of carbon dots via hydrothermal microwave treatment. *Inorganic Chemistry Communications*, 134, 109021.
23. Heer, E., Ruan, Y., Mealey, N., Quan, M. L., & Brenner, D. R. (2020). The incidence of breast cancer in Canada 1971–2015: trends in screening-eligible and young-onset age groups. *Canadian Journal of Public Health*, 111, 787-793.
24. Hofmann, M., Tauber, C., Hensel, N., & Thimme, R. (2021). CD8+ T cell responses during HCV infection and HCC. *Journal of Clinical Medicine*, 10(5), 991.
25. Hu, X., Zhang, Q., Xing, W., & Wang, W. (2022). Role of microRNA/lncRNA intertwined with the wnt/ β -catenin Axis in regulating the pathogenesis of triple-negative breast cancer. *Frontiers in pharmacology*, 13, 814971.

26. Iatrakis, G., & Zervoudis, S. (2021). Epidemiology of ductal carcinoma in situ. *Chirurgia (Romania)*, 116(5 Suppl), S15-S21.
27. Jagsi, R., Mason, G., Overmoyer, B. A., Woodward, W. A., Badve, S., Schneider, R. J., ... & Susan G. Komen-IBCRF IBC Collaborative in partnership with the Milburn Foundation. (2022). Inflammatory breast cancer defined: proposed common diagnostic criteria to guide treatment and research. *Breast Cancer Research and Treatment*, 192(2), 235-243.
28. Janardhan, B., Vaderhobli, S., Bhagat, R., Chennagiri Srinivasamurthy, P., Venketeshiah Reddihalli, P., Gawari, R., & Krishnamoorthy, L. (2015). Investigating impact of vascular endothelial growth factor polymorphisms in epithelial ovarian cancers: a study in the Indian population. *PloS one*, 10(7), e0131190.
29. Kazeminava, F., Javanbakht, S., Latifi, Z., Rasoulzadehzali, M., Abbaszadeh, M., Alimohammadzadeh, B., ... & Nouri, M. (2024). Ultrasound-assisted encapsulating folic acid-based carbon quantum dots within breast cancer cell-derived exosomes as a co-receptors-mediated anticancer nanocarrier for enhanced breast cancer therapy. *Scientific Reports*, 14(1), 16941.
30. Kendall, T., Verheij, J., Gaudio, E., Evert, M., Guido, M., Goepfert, B., & Carpino, G. (2019). Anatomical, histomorphological and molecular classification of cholangiocarcinoma. *Liver International*, 39, 7-18.
31. Khoshnood, A., Farhadian, N., Abnous, K., Matin, M. M., Ziaee, N., & Yaghoobi, E. (2023). N doped-carbon quantum dots with ultra-high quantum yield photoluminescent property conjugated with folic acid for targeted drug delivery and bioimaging applications. *Journal of Photochemistry and Photobiology A: Chemistry*, 444, 114972.
32. Köbel, M., & Kang, E. Y. (2022). The evolution of ovarian carcinoma subclassification. *Cancers*, 14(2), 416.

- 33.Lim, C. J., Lee, Y. H., Pan, L., Lai, L., Chua, C., Wasser, M., ... & Chew, V. (2019). Multidimensional analyses reveal distinct immune microenvironment in hepatitis B virus-related hepatocellular carcinoma. *Gut*, *68*(5), 916-927.
- 34.Liu, Z. C., Liu, W. D., Liu, Y. H., Ye, X. H., & Chen, S. D. (2015). Multiple sexual partners as a potential independent risk factor for cervical cancer: a meta-analysis of epidemiological studies. *Asian Pacific Journal of Cancer Prevention*, *16*(9), 3893-3900.
- 35.Llovet, J. M., Willoughby, C. E., Singal, A. G., Greten, T. F., Heikenwälder, M., El-Serag, H. B., ... & Friedman, S. L. (2023). Nonalcoholic steatohepatitis-related hepatocellular carcinoma: pathogenesis and treatment. *Nature reviews Gastroenterology & hepatology*, *20*(8), 487-503.
- 36.Luo, L., Song, T., Wang, H., Yuan, Q., & Zhou, S. (2018). A highly selective fluorescence sensing platform for nanomolar Hg (II) detection based on cytosine derived quantum dot. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, *193*, 95-101.
- 37.Ma, C., Kesarwala, A. H., Eggert, T., Medina-Echeverz, J., Kleiner, D. E., Jin, P., ... & Greten, T. F. (2016). NAFLD causes selective CD4⁺ T lymphocyte loss and promotes hepatocarcinogenesis. *Nature*, *531*(7593), 253-257.
- 38.Ma, L., Hernandez, M. O., Zhao, Y., Mehta, M., Tran, B., Kelly, M., ... & Wang, X. W. (2019). Tumor cell biodiversity drives microenvironmental reprogramming in liver cancer. *Cancer cell*, *36*(4), 418-430.
- 39.Mahdavi, M., Nassiri, M., Kooshyar, M. M., Vakili-Azghandi, M., Avan, A., Sandry, R., ... & Gopalan, V. (2019). Hereditary breast cancer; Genetic penetrance and current status with BRCA. *Journal of cellular physiology*, *234*(5), 5741-5750.
- 40.Maio, F., Tari, D. U., Granata, V., Fusco, R., Grassi, R., Petrillo, A., & Pinto, F. (2021). Breast cancer screening during COVID-19 emergency: patients and

- department management in a local experience. *Journal of Personalized Medicine*, 11(5), 380.
41. Mattiuzzi, C., & Lippi, G. (2020). Cancer statistics: a comparison between world health organization (WHO) and global burden of disease (GBD). *European journal of public health*, 30(5), 1026-1027.
42. Naik, G. G., Shah, J., Balasubramaniam, A. K., & Sahu, A. N. (2021). Applications of natural product-derived carbon dots in cancer biology. *Nanomedicine*, 16(7), 587-608.
43. Nakamura, H., Arai, Y., Totoki, Y., Shiota, T., Elzawahry, A., Kato, M., ... & Shibata, T. (2015). Genomic spectra of biliary tract cancer. *Nature genetics*, 47(9), 1003-1010.
44. Obeagu, E. I., Ahmed, Y. A., Obeagu, G. U., Bunu, U. O., Ugwu, O. P. C., & Alum, E. U. (2023). Biomarkers of breast cancer: Overview. *Int. J. Curr. Res. Biol. Med*, 1, 8-16.
45. Pallett, L. J., Gill, U. S., Quaglia, A., Sinclair, L. V., Jover-Cobos, M., Schurich, A., ... & Maini, M. K. (2015). Metabolic regulation of hepatitis B immunopathology by myeloid-derived suppressor cells. *Nature medicine*, 21(6), 591-600.
46. Park, H. R., Park, S. B., Hong, H. D., Suh, H. J., & Shin, K. S. (2017). Structural elucidation of anti-metastatic rhamnogalacturonan II from the pectinase digest of citrus peels (*Citrus unshiu*). *International journal of biological macromolecules*, 94, 161-169.
47. Perkins, R. B., Wentzensen, N., Guido, R. S., & Schiffman, M. (2023). Cervical cancer screening: a review. *Jama*, 330(6), 547-558.
48. Pfister, D., Núñez, N. G., Pinyol, R., Govaere, O., Pinter, M., Szydlowska, M., ... & Heikenwalder, M. (2021). NASH limits anti-tumour surveillance in immunotherapy-treated HCC. *Nature*, 592(7854), 450-456.

49. Poursadegh, H., Amini-Fazl, M. S., Javanbakht, S., & Kazeminava, F. (2024). Magnetic nanocomposite through coating mannose-functionalized metal-organic framework with biopolymeric pectin hydrogel beads: A potential targeted anticancer oral delivery system. *International Journal of Biological Macromolecules*, 254, 127702.
50. Prabhu, M., & Eckert, L. O. (2016). Development of World Health Organization (WHO) recommendations for appropriate clinical trial endpoints for next-generation Human Papillomavirus (HPV) vaccines. *Papillomavirus Research*, 2, 185-189.
51. Prieto-Montero, R., Katsumiti, A., Cajaraville, M. P., López-Arbeloa, I., & Martínez-Martínez, V. (2020). Functionalized fluorescent silica nanoparticles for bioimaging of cancer cells. *Sensors*, 20(19), 5590.
52. Qiao, L., Sun, T., Zheng, X., Zheng, M., & Xie, Z. (2018). Exploring the optimal ratio of d-glucose/l-aspartic acid for targeting carbon dots toward brain tumor cells. *Materials Science and Engineering: C*, 85, 1-6.
53. Rice, M. S., Poole, E. M., Willett, W. C., & Tworoger, S. S. (2020). Adult dietary fat intake and ovarian cancer risk. *International journal of cancer*, 146(10), 2756-2772.
54. Sahai, E., Astsaturov, I., Cukierman, E., DeNardo, D. G., Egeblad, M., Evans, R. M., ... & Werb, Z. (2020). A framework for advancing our understanding of cancer-associated fibroblasts. *Nature Reviews Cancer*, 20(3), 174-186.
55. Sattariazar, S., Ebrahimi, S. N., Aarsalani, N., & Kazeminava, F. (2023). Encapsulation of thymol and menthol loaded N/S co-doped carbon dots derived from a mixture of herbal extracts as theranostic agents with anticancer properties. *Colloids and Surfaces B: Biointerfaces*, 232, 113603.
56. Saw, P. E., & Song, E. W. (2019). Phage display screening of therapeutic peptide for cancer targeting and therapy. *Protein & cell*, 10(11), 787-807.

57. Seong, B. K. A., Lau, J., Adderley, T., Kee, L., Chaukos, D., Pienkowska, M., ... & Irwin, M. S. (2015). SATB2 enhances migration and invasion in osteosarcoma by regulating genes involved in cytoskeletal organization. *Oncogene*, *34*(27), 3582-3592.
58. Shereema, R. M., Sruthi, T. V., Kumar, V. S., Rao, T. P., & Shankar, S. S. (2015). Angiogenic profiling of synthesized carbon quantum dots. *Biochemistry*, *54*(41), 6352-6356.
59. Shi, C., Qi, H., Ma, R., Sun, Z., Xiao, L., Wei, G., ... & Guo, Z. (2019). N, S-self-doped carbon quantum dots from fungus fibers for sensing tetracyclines and for bioimaging cancer cells. *Materials Science and Engineering: C*, *105*, 110132.
60. Sia, D., Losic, B., Moeini, A., Cabellos, L., Hao, K., Revill, K., ... & Llovet, J. M. (2015). Massive parallel sequencing uncovers actionable FGFR2–PPHLN1 fusion and ARAF mutations in intrahepatic cholangiocarcinoma. *Nature communications*, *6*(1), 6087.
61. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. *CA Cancer J Clin* 2020; *70*: 7-30.
62. Singh, I., Arora, R., Dhiman, H., & Pahwa, R. (2018). Carbon quantum dots: Synthesis, characterization and biomedical applications. *Turk. J. Pharm. Sci*, *15*(2), 219-230.
63. Stein, C., Bardet, A. F., Roma, G., Bergling, S., Clay, I., Ruchti, A., ... & Bauer, A. (2015). YAP1 exerts its transcriptional control via TEAD-mediated activation of enhancers. *PLoS genetics*, *11*(8), e1005465.
64. Stelzle, D., Tanaka, L. F., Lee, K. K., Khalil, A. I., Baussano, I., Shah, A. S., ... & Dalal, S. (2021). Estimates of the global burden of cervical cancer associated with HIV. *The lancet global health*, *9*(2), e161-e169.

65. Su, W., Guo, R., Yuan, F., Li, Y., Li, X., Zhang, Y., ... & Fan, L. (2020). Red-emissive carbon quantum dots for nuclear drug delivery in cancer stem cells. *The journal of physical chemistry letters*, *11*(4), 1357-1363.
66. Tan, A. T., Yang, N., Krishnamoorthy, T. L., Oei, V., Chua, A., Zhao, X., ... & Bertoletti, A. (2019). Use of expression profiles of HBV-DNA integrated into genomes of hepatocellular carcinoma cells to select T cells for immunotherapy. *Gastroenterology*, *156*(6), 1862-1876.
67. Tan, M., Li, X., Wu, H., Wang, B., & Wu, J. (2015). N-doped carbon dots derived from bovine serum albumin and formic acid with one-and two-photon fluorescence for live cell nuclear imaging. *Colloids and Surfaces B: Biointerfaces*, *136*, 141-149.
68. Timmermans, M., Sonke, G. S., Van de Vijver, K. K., Van der Aa, M. A., & Kruitwagen, R. F. P. M. (2018). No improvement in long-term survival for epithelial ovarian cancer patients: A population-based study between 1989 and 2014 in the Netherlands. *European journal of cancer*, *88*, 31-37.
69. Tiwari, P., Shukla, R. P., Yadav, K., Singh, N., Marwaha, D., Gautam, S., ... & Mishra, P. R. (2024). Dacarbazine-primed carbon quantum dots coated with breast cancer cell-derived exosomes for improved breast cancer therapy. *Journal of Controlled Release*, *365*, 43-59.
70. Tiwari, P., Yadav, K., Shukla, R. P., Gautam, S., Marwaha, D., Sharma, M., & Mishra, P. R. (2023). Surface modification strategies in translocating nanovesicles across different barriers and the role of bio-vesicles in improving anticancer therapy. *Journal of Controlled Release*, *363*, 290-348.
71. Unnikrishnan, B., Wu, R. S., Wei, S. C., Huang, C. C., & Chang, H. T. (2020). Fluorescent carbon dots for selective labeling of subcellular organelles. *Acs Omega*, *5*(20), 11248-11261.

72. Vatankhah, H., Khalili, P., Vatanparast, M., Ayoobi, F., Esmaeili-Nadimi, A., & Jamali, Z. (2023). Prevalence of early and late menopause and its determinants in Rafsanjan cohort study. *Scientific Reports*, *13*(1), 1847.
73. Vijgen, S., Terris, B., & Rubbia-Brandt, L. (2017). Pathology of intrahepatic cholangiocarcinoma. *Hepatobiliary surgery and nutrition*, *6*(1), 22.
74. Wang, J., Chen, D., Li, B., He, J., Duan, D., Shao, D., & Nie, M. (2016). Fe-MIL-101 exhibits selective cytotoxicity and inhibition of angiogenesis in ovarian cancer cells via downregulation of MMP. *Scientific Reports*, *6*(1), 26126.
75. Wang, Y., Chen, J., Tian, J., Wang, G., Luo, W., Huang, Z., ... & Fan, X. (2022). Tryptophan-sorbitol based carbon quantum dots for theranostics against hepatocellular carcinoma. *Journal of nanobiotechnology*, *20*(1), 78.
76. Wu, H., Su, W., Xu, H., Zhang, Y., Li, Y., Li, X., & Fan, L. (2021). Applications of carbon dots on tumour theranostics. *View*, *2*(2), 20200061.
77. Xia, C., Dong, X., Li, H., Cao, M., Sun, D., He, S., ... & Chen, W. (2022). Cancer statistics in China and United States, 2022: profiles, trends, and determinants. *Chinese medical journal*, *135*(05), 584-590.
78. Yang, J., Wang, Q., Xing, T., Wang, X., Li, G., Shang, Z., ... & Ou, T. (2023). Engineered exosome-mediated cobalt sulfide quantum dot targeted delivery for photothermal and chemodynamic anticancer therapy. *Journal of Drug Delivery Science and Technology*, *83*, 104441.
79. Yuan, Y., Cai, X., Shen, F., & Ma, F. (2021). HPV post-infection microenvironment and cervical cancer. *Cancer Letters*, *497*, 243-254.
80. Zahed, Z., Hadi, R., Imanzadeh, G., Ahmadian, Z., Shafiei, S., Zadeh, A. Z., ... & Kazeminava, F. (2024). Recent advances in fluorescence nanoparticles “quantum dots” as gene delivery system: A review. *International Journal of Biological Macromolecules*, *254*, 127802.

- 81.Zhang, S., Xu, H., Zhang, L., & Qiao, Y. (2020). Cervical cancer: Epidemiology, risk factors and screening. *Chinese Journal of Cancer Research*, 32(6), 720.
- 82.Zhou, L., Yao, L., Dai, L., Zhu, H., Ye, X., Wang, S., ... & Chang, X. (2021). Ovarian endometrioid carcinoma and clear cell carcinoma: A 21-year retrospective study. *Journal of ovarian research*, 14, 1-12.
- 83.Zhu, Y. S., Tang, K., & Lv, J. (2021). Peptide–drug conjugate-based novel molecular drug delivery system in cancer. *Trends in pharmacological sciences*, 42(10), 857-869.
- 84.Zuber, M., Chhabra, M., Venkataraman, R., Kumar, S., & Rashid, M. (2022). Methotrexate related cutaneous adverse drug reactions: A systematic literature review. *Journal of Basic and Clinical Physiology and Pharmacology*, 33(5), 549-565.